Renal Transplantation

- 1933 First cadaveric renal allograft Voronay in Ukraine 48 hr survival
- 1951 Azathioprine available for human use
- 1955 First successful Tx intertwin renal transfer, Murray 1955 (Boston US)
- 1962 Corticosteroids with azathioprine
- 1967 Renal preservation media and pulse preservation
- 1966 Direct crossmatch introduced
- 1970s Brain death laws in US and beating heart cadaveric harvesting
- 1978 Cyclosporin introduced
- 1980s University of Wisconsin media

Outcomes

| | Live donor | Cadaveric donor |
|--------------------|------------|-----------------|
| Patient survival | | |
| 1 year | 97.8% | 95% |
| 3 year | 95.2% | 89.2% |
| 5 year | 90.5% | 81.3% |
| Allograft survival | | |
| 1 year | 94.7% | 89.2% |
| 3 year | 87.4% | 77.7% |
| 5 year | 76.0% | 61.3% |

Adverse risk factors

Extremes of recipient age (> 55yrs or < 1 yr)

Extremes of donor age (> 50 yrs or < 2 yrs)

Previous failed transplant

BMI > 30

Diabetes

Blacks

Coronary artery disease

Hypercoagulability

Risk factors for recurrent renal disease (#1 diabetes)*

* Systemic or primary renal disease

Systemic Diabetes, HUS, HSP, SLE, cryoglobulins, Wegener's,

scleroderma, sickle cell, oxalosis

Renal IgA, membranoproliferative, anti-GBM, focal segmental

glomerulosclerosis

NB. HUS, focal segmental glomerulosclerosis and primary oxalosis very high risk of recurrence and graft failure

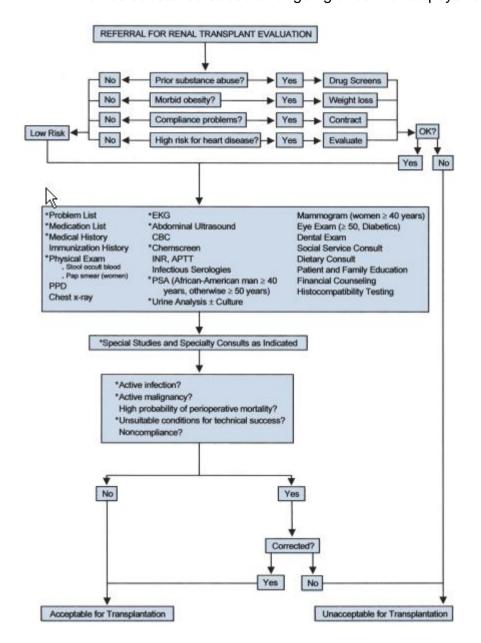
Recipient evaluation

Confirm irreversible renal failure

Exclude significant systeic medical disease (including infection)

Exclude active malignancy or treated cancer with high risk of relapse

Exclude substance abuse or ongoing uncontrolled psychiatric disease



Serological testing: HBV, HCV, CMV, syphillis, EBV, HSV, HIV, HTLV-1,

Toxoplasma, TB, VZV

Vaccination: Pneumococcus (if < 6yrs)

Influenza

Hepatitis B (0, 1 and 6 months pre-op)

Varicella zoster (if seronegative)

Tetanus diptheria toxoid

Polio booster

MMR

Cancer: Disease-free intervals of 2-5 years recommended

Shorter for individual cancers? low risk RCC

No wait for skin cancers and CIS cervix or primary brain

tumours

Cholecystectomy for gallbladder polyps > 1cm

Urological: History to exclude dysfunctional voiding

Urinalysis to exclude infection/microhaematuria

USS and postvoid residual to exclude hydro and poor

emptying

TURP/BNI may be required

Diversion not necessarily a contraindication: transplant

into pouch/conduit reportedly succesful

Biopsy to exclude fibrosis +/- wet augment recommended for patients diverted for reflux – capacity rapidly returns to normal in absence of fibrosis (pre-Tx Nx usually required)

Other: ADPKD = cerebral aneurysm screening

Peripheral arterial or venous disease = dopplers

Indications for pre-transplant nephrectomy (Campbells)

Renal stones not cleared by minimally invasive techniques or lithotripsy

Solid renal tumors with or without acquired renal cystic disease

Polycystic kidneys that are symptomatic, extend below the iliac crest, have been infected, or have solid tumors

Persistent antiglomerular basement membrane antibody levels

Significant proteinuria not controlled with medical nephrectomy or angioablation

Recurrent pyelonephritis

Grade 4 or 5 hydronephrosis

S Stone

T Tumour

I Infection

M Massive

P Proteinuria

4 Grade 4/5 reflux

Contraindications to renal transplantation

Active systemic renal disease (SLE, anti-GBM, HUS, ANCA + GN)

Oxalosis (combined renal and liver transplant)

Active infection

Recently treated or uncontrolled/disseminated malignancy

Prohibitive extrarenal disease (CVS etc.)

Active IV or alcohol abuse

Non-compliance

Uncontrolled psychiatric disorder

M Malignancy

I Infection

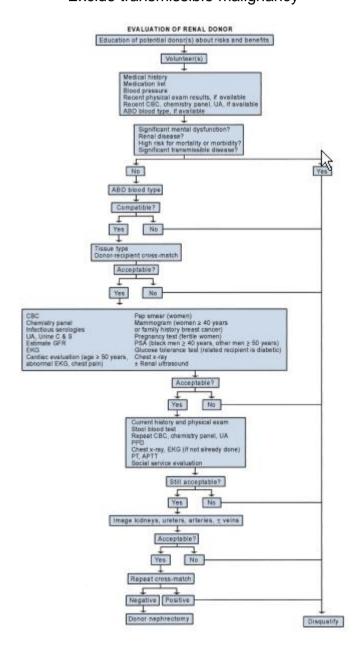
N Nephropathy

C Compliance issues

- E Severe extrarenal disease
- S Substance abuse
- O Oxalosis
- P Psychiatric disorder

Living donor evaluation

Exclude renal disease
Exclude active infection
Exclde transmissible malignancy



Best kidney left behind: ~ 80% CrCl achieved post-op – hyperfiltration leads to IL-10 production and TGF-B release.

If both equal choose right side in female of childbearing age

Imaging of choice = 3D CT angiography with delayed images best modality to image arteries, veins and drainage

Most living donor allografts now harvested laparoscopically - cuff of IVC on R

Typically mannitol given prior to renal vessel clamping (diuresis/free-radicals) Overall:

Morbidity 1.8-7% general complications (cardiac, thrombotic, lungs)

Slightly increased risk of HT and proteinuria vs. general

population but clinically insignificant

0.1% risk of ESRF long-term

Mortality 0.02%

Cadaveric donors

Beating heart donors with clinically determined brain stem death Similar criteria to above re. malignancy, active infection etc.

Age 6 – 50 yrs acceptable

Low numbers resulted in 'extended criteria', but graft survival poorer in those younger than 6 or older than 50. Young donor kidneys can be used as pairs 'en bloc' with attached IVC & aorta. Also Light criteria utilising biopsy findings and clinical criteria used to predict risk of transplant failure Always go for left-side if possible and single renal vessels.

Organ preservation after procurement

Warm ischaemia – failure of oxidative phosphorylation - ATP depletion – Na/K pump failure – intracellular Na accumulation – cell swelling – no-reflow after revascularisation. Also free-radical production via hypoxanthine Cold ischaemia reduces energy requirements dramatically

2 methods: Simple cold slush storage (UoW)

Pulsatile perfusion

(i) Cold slush storage

Good kidneys

Storage up to 48 hours at 4'C

Cold ischaemia times > 24 hours a/w higher incidence of delayed graft function. Recommendation <21 hours.

University of Wisconsin solution – better vs. Eurocollins (Ploeg 1992) UoW consituents

- a. Lactobionate, starch, raffinose HMW solutes limit sweling
- b. Phosphate buffer H+ ions
- c. Adenosine substrate for ATP synthesis
- d. Glutathione free-radical scavenger
- e. Allopurinol inhibits xanthine oxidase and free-radicals
- f. Mg and DXM membrane stabilisers
- g. Penicillin antibiosis
- h. Insulin

(ii) Pulsatile perfusion

Poor kidneys (elderly donors, marginal criteria)

Allows assessment of flow (> 100ml/min), pressure (<40/25 mmg) and resistance (<0.3) features required for Tx

Transplantation – operative technique

Bench table vascular reconstruction important in cadaveric and LD Nx lap – removal of staple lines and construction of single venous and arterial channels if necessary

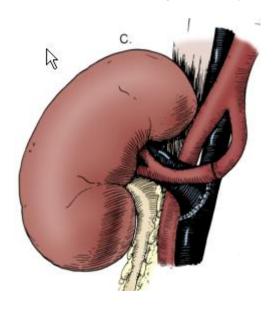
cadaver – use of IVC to lengthen renal vein on right. Construction of single vessel channels

Give antibiotics, steroids and immunoglobulins at induction

Typically RIF – external iliac vein more superficial cf. left

Extraperitoneal approach favoured in UK (avoids intestinal complications, ease of biopsy, confines surgical mischief)

Rutherford-Morrison type incision pubic symphysis to point 4 fingers above ASIS. Preservation of vas, division of round ligament and inferior epigastric. Mobilisation of peritoneum medially. Heparin given before vessel clamping. mannitol at time of anastomosis. End-to-end arterial anastomosis first from renal artery to either hypogastric artery or end-to-side to external iliac artery. Then venous anastomosis external iliac (arterial anastomosis more critical and reduces external iliac clamping time). Lastly extravesical antirefluxing ureteroneocystostomy (transvesical or extravesical) – Lich Gregoir in UK vs. Leadbetter-Politano in US (higher incidence of reflux in UK but lower incidence of bladder dysfunction).



Post-operative complications

Early and late transplant-related complications and long-term non-surgical complications of renal transplantation.

Surgical complications causing allograft dysfunction divided into renal, vascular or urological complications:

A. Early allograft dysfunction

Renal

Acute tubular necrosis*
Prolonged cold ischaemia time and reperfusion injury*
Acute rejection (see below)
Acute cyclosporin toxicity

Vascular

Venous thrombosis

Arterial occlusion

Haemorrhage

AV fistula

Urological

Urine leak or fistula Urinary obstruction

* Leads to delayed graft function, defined as the need for dialysis in the first week after Tx

The above are investigated by allograft USS with Doppler, cyclosporin levels and if necessary, renal biopsy

B. Late allograft dysfunction

Renal

Chronic rejection

Chronic cyclosporin toxicity

Vascular

Renal artery stenosis

Urological

Urinary tract obstruction

UTI

Urinary stones

C. Medical consequences of renal transplantation

Coronary artery disease

Diabetes

Malignancy

Reproductive

Allograft rejection

Hyperacute, accelerated, acute and chronic:

(i) Hyperacute

Very rare

Immediate to 24 hours post-Tx

Pre-formed cytotoxic antibodies to allograft MHC – should not occur in absence of negative compatability screening

No Rx – nephrectomy required

(ii) Accelerated

Rare

Up to 10 days post-Tx

Primary immune response – humoral and cell-mediated

Very aggressive and difficult to treat – may result in graft rupture

Less common with anti-lymphocyte induction therapy

(iii) Acute

Common – up to 50% of transplants

Up to 6 months post-Tx

Raised serum creatinine +/- systemic flu-like symptoms, graft tenderness and hypertension

Cause unknown – diagnosed on biopsy after exclusion of infection

Typically lymphocytic infiltrate around tubules and vessels - severity graded by Banff criteria

Usually amenable to corticosteroid anti-rejection Rx (+ antilymphocyte agents if no response at 48 hrs)

(iv) Chronic

Common

Used to be called chronic allograft rejection – now IFTA (interstitial fibrosis and tubular atrophy)

Progressive decline in renal function leading to failure

Thought due to repeated immunological insults/toxicity due to immunosuppressive medications

Typically intimal proliferation and interstitial fibrosis – relative absence of lymphocytic infiltrate cf. acute rejection (can be difficult to differentiate from chronic cysclosporin toxicity)

No specific Rx – limitation of other insults (DM etc.)

Vascular complications

Renal artery stenosis

1-15% incidence

Due to surgeon error (narrowing, kink), arteriosclerosis, fibrosis

Typically presents with poorly controlled HT +/- bruit

> 70% stenosis >15mmHg gradient

Suggested by Doppler, ACEI renography, angiography (best)

Rx = angioplasty: outcome for surgery poor

Renal artery thrombosis

Rare < 5%

Surgeon error – intimal flap or kink

Diagnosed on Doppler (no flow) or renogram (photopenia)

Rarely salvageable - Nx

Renal vein thrombosis

Rare

Technical error, haematoma, iliofemoral DVT, pro-thrombotic state, Diagnosed on renogram (background scintillations) or Doppler (reversal of venous flow)

Rx = Thrombolysis/thrombectomy, Nx common

Urological complications

Infection

Stones

Urinary obstruction (3%)

Early (haematoma, urinoma, lymphocoele, surgeon error) or late (ischaemia or fibrosis, stone, tumour, bladder pathology)

Non-contrast imaging and standard endourological Mx

Most ureteric strictures due to ischaemia – exacerbated by BK virus Fistula (2-3%)

Usually due to devascularisation, ischaemia and necrosis

Typically distal third ureter (all supply from allograft renal artery)

Rx small leaks conservative with stent Rx large leaks/ necrosis reconstruction options:

Allograft – native ureteroureterostomy

Allograft – native pyeloureterostomy Allograft – native calicoureterostomy Calicocystostomy Boari flap +/- psoas hitch Ileal interposition

Lymphocoele

0.5 - 20% incidence

Asymptomatic; occasionally graft dysfunction or LUTS

Often recur following aspiration – open or lap marsupialisation vs. aspiration and cavity sclerosis

Medical consequences

Post-transplant diabetes

Incidence 3.4 - 46%

Generally due to corticosteroids; also reported with cyclosporin and tacrolimus; often settle

3 wks to 19 yrs post-transplant

Non-insulin or insulin-dependent

Worse patient and graft survival

Malignancy

Post-transplant lymphoproliferative disease*

~ 2-3 yrs ~ 2-3 yrs

Kaposi's sarcoma

Squamous carcinomas (skin, cervix, vulva and anus) ~8-10 yrs All above occur with greater frequency in transplant popn. Due to

combination immunosuppression

No difference for lung, breast, prostate, colon or uterine, but earlier onset and more aggressive

PSA levels normal in renal failure

Cytology reliable in transplant patients

BCG OK but increases risk of sepsis

Urothelial cancer risk slightly increased due to BK virus (decoy cells on cytology, diagnosed with blood and urine PCR – Rx reduce immunosuppression, Rx urothelial tumour)

*PTLD ~1% of renal transplants

Not the same as NHL A/w Epstein Barr virus Aggressive vs. normal Multicentric and extra-nodal

CNS and allograft involvement relatively common

*EBV infection

Increased risk of post-transplant lymphoproliferative disease

EBV viraemia on PCR

Reduce immunosuppression and give Rituximab (if CD25 positive)

CMV infection

Lethargy, respiratory symptoms and hepatitis

Destauration on mobilisation

Ground glass appearance on CXR

Serology and PCR reaction for diagnosis

Screen donor and recipient

3 months prophylaxis with septrin

Valgancicylovir and for Rx; foscarnet for resistance Reproductive

Male infertility

Sperm parameters improve following Tx but not to normal levels – many cases of pregnancy reported

Serum T and FSH/LH usually normalise following Tx suggesting improved Leydig cell function in non-uraemic state

Pregnancy

Possible but increased risks of pre-eclampsia (25%), pre-term labour (~50%) and neonatal morbidity (30%)
Graft function deteriorates in pregnancy, with 5-10% cases developing permanent graft dysfunction
No evidence that graft position impairs foetal delivery
Chromosomal aberrations higher in offspring but no definite increase in haematological malignancy

Attempts at pregnancy generally allowed after ~12 months

Appendix

<u>Histocompatability</u>

ABO blood groups and major histocompatability complex (MHC)

(i) ABO blood groups

Blood antigens (eg. A and B) present on endothelial cells in transplant kidney – recipient blood contains antibodies to antigen they lack

Cross match crucial to assess compatibility

Occasionally A2 kidneys transplanted into O or B donors with low anti-A2 antibodies

(ii) MHC antigens

Encoded by MHC genes on short arm chromosome 6 Glycoproteins on cell membrane of all cells:

HLA-A, HLA-B. HLA-C

Bind 8-10 amino acids in groove

Interacts with CD8

Subtype detected by tissue typing T

lymphocytes

Class 2 MHC antigens B and activated T lymphocytes and

antigen presenting cells HLA-DR, HLA-DQ, HLA-DP

Bind 12-28 amino acids in groove

Interacts with CD4

Subtype detected by tissue typing B

lymphocytes

Huge variation in HLA/MHC gene polymorphisms (see below) allows humans as a species to respond to a myriad of potential pathogens

| HLA SPECIFICITIES | | | | | | |
|-------------------|---------|---------|----------|---------|--------|------|
| A | В | В | С | DR | DQ | DP |
| A1 | B5 | B51(5) | Cw1 | DR1 | DQ1 | DPw1 |
| A2 | B7 | B5102 | Cw2 | DR103 | DQ2 | DPw2 |
| A203 | B703 | B5103 | Cw3 | DR2 | DQ3 | DPw3 |
| A210 | B8 | B52(5) | Cw4 | DR3 | DQ4 | DPw4 |
| A3 | B12 | B53 | Cw5 | DR4 | DQ5(1) | DPw5 |
| A9 | B13 | B54(22) | Cw6 | DR5 | DQ6(1) | DPw6 |
| A10 | B14 | B55(22) | Cw7 | DR6 | DQ7(3) | |
| A11 | B15 | B56(22) | Cw8 | DR7 | DQ8(3) | |
| A19 | B16 | B57(17) | Cw9(w3) | DR8 | DQ9(3) | |
| A23(9) | B17 | B58(17) | Cw10(w3) | DR9 | | |
| A24(9) | B18 | B59 | | DR10 | | |
| A2403 | B21 | B60(40) | | DR11(5) | | |
| A25(10) | B22 | B61(40) | | DR12(5) | | |
| A26(10) | B27 | B62(15) | | DR13(6) | | |
| A28 | B2708 | B63(15) | | DR14(6) | | |
| A29(19) | B35 | B64(14) | | DR1403 | | |
| A30(19) | B37 | B65(14) | | DR1404 | | |
| A31(19) | B38(16) | B67 | | DR15(2) | | |
| A32(19) | B39(16) | B70 | | DR16(2) | | |
| A33(19) | B3901 | B71(70) | | DR17(3) | | |
| A34(10) | B3902 | B72(70) | | DR18(3) | | |
| A36 | B40 | B73 | | DR51 | | |
| A43 | B4005 | B75(15) | | DR52 | | |
| A66(10) | B41 | B76(15) | | DR53 | | |
| A68(28) | B42 | B77(15) | | | | |
| A69(28) | B44(12) | B7801 | | | | |
| A74(19) | B45(12) | B81 | | | | |
| A80 | B46 | Bw4 | | | | |
| | B47 | Bw6 | | | | |
| | B48 | | | | | |
| | B49(21) | | | | | |
| | B50(21) | | | | | |

However, makes life difficult when trying to find compatible matches

Detecting incompatability

Originally complement dependent cytotoxicity (CDC) used, utilising donor lymphocytes and recipient sera in direct tests, with rabbit complement to stimulate cell lysis. However dependent on complement pathways and not that sensitive or specific.

Flow cytometry or ELISA based tests (known as panel reactivity antibody testing (PRA)) have now become standard to give an indication of the likelihood of chronic rejection. However ultimate decision to use kidney based on HLA compatibility. In general matching at only 3 loci; HLA-A, HLA-B and HLA-DR:

000 full house match

222 complete mismatch

Low risk 000 or single mismatch at non-DR locus

Medium risk 2B mismatches or 1DR mismatch High risk 2DR mismatch, 2nd transplant

Life of transplant related to degree of match:

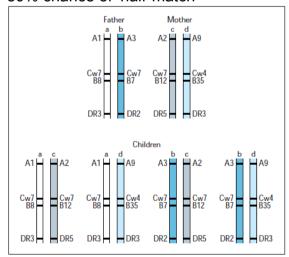
| HLA Antigens Mismatched | No. | Half-Life (Yr)[*] |
|--------------------------------|--------|-------------------|
| 0 | 4,182 | 15 |
| 1–3 | 9,391 | 12 |
| 4–6 | 14,186 | 10 |

Before transplantation a manual XM is performed between recipient serum and donor lymphocytes in all cases to exclude hyperacute rejection

Living-related donors (see below)

Each individual has 2 x chromosome 6 Parent shares 50% chromosomes with child Siblings

25% chance of perfect match or mismatch 50% chance of 'half-match'



Immunology of graft rejection

Recognition of allograft as foreign occurs in two ways: direct recognition of donor APCs and recipient processing of donor peptides by host APCs (indirect recognition)

Activation of T-cells requires 3 signals: anigen presentation, glycoprotein binding (CD28 etc.) and IL-2. IL-2 require to provide co-stimulatory signal to drive cytotoxicity. Killing mediated in three ways: cytotoxic T-lymphocytes, antibody mediated (complement activation, opsonisation), delayed type hypersensitivity

<u>Immunosuppresive therapy and side effects</u>

A. Induction therapy designed to prevent rejection in first 3 months

IL-2 receptor antagonist (basiliximab) anti B-cell Anti-thymocyte globulin anti T-cell*

Adeluzumab anti B and T-cell

* very toxic and requires central line - only for high risk induction

B. Chronic anti-rejection therapy

Typically triple therapy:

(i) calcineurin inhibitor

cyclosporine, tacrolimus

(ii) purine inhibitor

azathiaprine, mycophenolate mofetil

(iii) steroids

Combination of above depends on risk of rejection (see above) – each nephrology unit has own protocols. Typically:

Low risk cyclosporine, azathiaprine, steroid

High risk tacrolimus, mycophenylate mofetil, steroid

Recently rapamycin used alone or in combination with cyclosporine in an attempt to reduce steroid intake

Pulsed steroids for acute rejection; anti-lymphocyte agents for non-responders

NB. Cyclosporin and tacrolimus metabolised by cytochrome p450 system – inhibitors increase dose (ABx except anti-TB drugs; Cachannel blockers); inducers reduced dose (anti-TB ABx; epilepsy drugs)

| Immunosuppressant | Mechanism of Action | Interferes with |
|-------------------------------|---|---|
| Glucocorticoids | Reduce transcription of cytokine genes | Intracellular signaling |
| Azathioprine | Inhibits purine synthesis | Lymphocyte proliferation |
| Mycophenolate mofetil | Inhibits purine synthesis | Lymphocyte proliferation |
| Sirolimus | Inhibits cell cycle progression | Lymphocyte proliferation |
| Tacrolimus | Inhibits calcineurin and interleukin-2 production | Intracellular signaling |
| Cyclosporine | Inhibits calcineurin and interleukin-2 production | Intracellular signaling |
| Monomurab CD3 | Depletes T lymphocytes | Antigen recognition |
| Equine antithymocyte globulin | Depletes T lymphocytes | Antigen recognition |
| Rabbit antithymocyte globulin | Depletes T lymphocytes | Antigen recognition |
| Alemtuzumab (off label) | Depletes T and B lymphocytes | Antigen recognition and antibody production |
| Rituximab (off label) | Depletes B lymphocytes | Antibody production |
| Basiliximab | Blocks interleukin-2 receptor | Intercellular signaling |
| Daclizumab | Blocks interleukin-2 receptor | Intercellular signaling |

Side Effects of Immunosuppressive Medications

Corticosteroids Azathioprine **Diabetes** Bone marrow suppression (pancytopenia) Lipid disorders Gastrointestinal disturbances Cushingoid features Hepatotoxicity Obesity Hair loss Poor wound healing Antilymphocytic agents Avascular necrosis (bone) Polyclonal Cataracts Fever, chills Hypertension, coronary artery disease (CAD) Leukopenia, thrombocytopenia Peptic ulceration, gastritis, bowel perforation Serum sickness Growth retardation Local phlebitis **Pancreatitis** Monoclonal Cyclosporine Flulike syndrome: Nephrotoxicity Fever, chills, tremors, headache, nausea, Hypertension, CAD vomiting, and diarrhea Hyperkalemia Aseptic meningitis Hyperuricemia Hyptotension Heptatotoxicity Pulmonary edema Hirsutism FK506 (Prograf/Tacrolimus) Gingival hyperplasia Nephrotoxicity Tremors/seizures **Diabetes Pancreatitis** Neurotoxicity

Hemolytic-uremic syndrome

GI bleeding, diarrhea, ulceration,

Mycophenolate Mofetil (CellCept)

Gastrointestinal disturbances

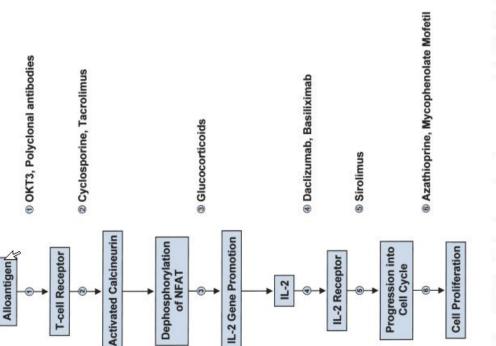
esophagitis, gastritis

Leukopenia

Simulect Rapamune (Rapamycin)

Interleukin Inhibitor

Bone marrow suppression GI toxicity Skin rash Hyperlipidemia



Basic immunology

<u>Innate</u>

NK cells

Complement

Acute phase proteins

Physical and mucosal barriers

Acquired

MHC class I antigens HLA A, B and C

All nucleated cells, including T-cells

Interact with CD8 on immune cells

MHC class II antigens HLA DR, DP, DQ

Antigen presenting cells

Macrophages Monocytes Some B cells

Langerhan's skin cells

Dendritic cells

Vascular endothelial cells Interact with CD4 receptor on T-cells

Antigen presentation Occurs in peripheral lymph nodes

APCs interact with T-cells

Immune activity vs. infections

Innate and acquired

Extracellular bacteria Complement killing

NK activity

Opsonisation and macrophage killing

Intracellular bacteria T-cell activation required

Immune activity vs. tumours

Tumours are antigenic, but evade immune mediated killing in a number of ways:

Downregulation in MHC class I and II

Decreased expression of transporter antigen proteins (TAPs)

Overexpression of TGF-B